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# KLMab-1: An Anti-human KLRG1 Monoclonal Antibody for Immunocytochemistry

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Immune checkpoint molecules have received attention as targets of cancer immunotherapy. Killer cell lectin-like receptor subfamily G member 1 (KLRG1) is one of the immune checkpoint molecules expressed in CD4<sup>+</sup> T, CD8<sup>+</sup> T, and natural killer (NK) cells. KLRG1 exhibits antiviral and antitumor immunity, and its expression in T and NK cells is upregulated by viral infectious diseases and some tumors. Thus, monoclonal antibodies (mAbs) for KLRG1 would be useful tools for the diagnosis and immunotherapy against viral infectious diseases and cancers. We have developed anti-human KLRG1 (hKLRG1) mAb (clone KLMab-1, mouse IgG<sub>1</sub>, kappa) by the Cell-Based Immunization and Screening method. We have also demonstrated that KLMab-1 recognizes both exogenous and endogenous hKLRG1 in flow cytometry. In this study, we first showed that KLMab-1 and its recombinant mAb (recKLMab-1) bound to exogenous hKLRG1 overexpressed in Chinese hamster ovary (CHO)-K1 cells, but not in parental CHO-K1 cells, in immunocytochemistry. We next showed that both mAbs detected endogenous hKLRG1 expressed in human NK cells. These results demonstrate that KLMab-1 and recKLMab-1 are available for immunocytochemistry.

**Keywords:** KLRG1, KLMab-1, monoclonal antibody, immunocytochemistry

## Introduction

**T** CELLS AND NATURAL KILLER (NK) cells play crucial roles in antiviral and antitumor immunity.<sup>1-4</sup> During the response to viral infection or cancer development, T cells are activated, expanded, and differentiated into effector and memory T cells. Activated NK cells migrate to the infected or tumor site, and the NK cells eliminate the target cells through the production of cytokines and exhibition of cytolytic activity. In contrast, the activities of T and NK cells are suppressed by immune checkpoint molecules, including programmed cell death 1 (PD-1) and cytotoxic T lymphocyte-associated antigen 4 (CTLA-4).<sup>5</sup>

Accordingly, immune checkpoint molecules have drawn attention as targets for cancer immunotherapy. In fact, specific monoclonal antibodies (mAbs) against PD-1 and CTLA-4 have provided great advances in the medical treatment of cancers.<sup>6</sup> However, developing novel mAbs against other

immune checkpoint molecules has been required because the number of patients who respond to anti-PD-1 or anti-CTLA-4 mAbs is limited.<sup>7</sup>

Killer cell lectin-like receptor subfamily G member 1 (KLRG1), a lectin-like type II transmembrane protein, is an immune checkpoint molecule expressed in CD4<sup>+</sup> T, CD8<sup>+</sup> T, and NK cells.<sup>8-10</sup> It harbors an immunoreceptor tyrosine-based inhibitory motif (ITIM) in its cytoplasmic region. Upon binding to KLRG1 ligands including E-cadherin, KLRG1 evokes inhibitory signaling through recruitment of Src homology 2 domain-containing inositol polyphosphate 5-phosphatase 1 (SHIP1) and Src homology region 2 domain-containing phosphatase 2 (SHP2) to ITIM. Then, KLRG1 attenuates interferon  $\gamma$  production in T and NK cells, and suppresses NK cell-mediated cytotoxicity.<sup>8,10-16</sup>

The mechanism contributes to the progression of viral infection and tumor by KLRG1. Moreover, the expression of KLRG1 is increased in NK cells of virus-infected mice and

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T cells of cervical and colorectal cancer patients.<sup>8,17,18</sup> These reports have suggested that KLRG1 can be a target molecule for the diagnosis and immunotherapy of viral infectious diseases and some cancers.

We have established mAbs against cell surface-expressing membrane proteins by the Cell-Based Immunization and Screening, including CCR3,<sup>19–21</sup> CCR8,<sup>22–25</sup> CCR9,<sup>26,27</sup> CD10,<sup>28,29</sup> CD19,<sup>30</sup> CD20,<sup>31,32</sup> CD44,<sup>33</sup> CD133,<sup>34</sup> EpCAM,<sup>35,36</sup> HER3,<sup>37</sup> PD-L1,<sup>38</sup> podoplanin,<sup>39–53</sup> TIGIT,<sup>54</sup> and TROP2.<sup>55,56</sup> We have also established an antihuman KLRG1 (hKLRG1) mAb (clone KLMab-1; mouse IgG<sub>1</sub>, kappa), which reacts to endogenous and exogenous hKLRG1 in flow cytometry.<sup>57</sup> In this study, we showed that KLMab-1 and its recombinant mAb (recKLMab-1) are available for immunocytochemistry against endogenous and exogenous hKLRG1.

## Materials and Methods

### Cell lines

Chinese hamster ovary (CHO)-K1 cells were obtained from the American Type Culture Collection (Manassas, VA). CHO-K1 cells overexpressed with human KLRG1 (CHO/hKLRG1) were established previously.<sup>57</sup> CHO-K1 and CHO/hKLRG1 were cultured in Roswell Park Memorial Institute 1640 medium (Nacalai Tesque, Inc., Kyoto, Japan), supplemented with 10% heat-inactivated fetal bovine serum (Thermo Fisher Scientific, Inc., Waltham, MA), 100 U/mL of penicillin, 100 µg/mL of streptomycin, and 0.25 µg/mL of amphotericin B (Nacalai Tesque, Inc.). The cells were maintained in a humidified atmosphere at 37°C and 5% carbon dioxide. Human NK cells (donor lot. 4022602, purity >70%) were purchased from Takara Bio (Shiga, Japan).

### Antibodies

The development of KLMab-1 was described in our previous report.<sup>57</sup> To generate a recombinant KLMab-1 (recKLMab-1), V<sub>H</sub> and C<sub>H</sub> cDNAs of KLMab-1 were subcloned into the pCAG-Neo vector (FUJIFILM Wako Pure Chemical Corporation, Osaka, Japan), and V<sub>L</sub> and C<sub>L</sub> cDNAs of KLMab-1 were subcloned into the pCAG-Ble vector (FUJIFILM Wako Pure Chemical Corporation). An anti-hKLRG1 mAb (clone SA231A2) was purchased from BioLegend (San Diego, CA). Alexa Fluor 488-conjugated anti-mouse IgG was purchased from Cell Signaling Technology, Inc. (Danvers, MA).

### Immunocytochemistry

For immunocytochemistry of CHO-K1 and CHO/hKLRG1 cells, the cells were attached to an acid-wash coverslip and were fixed with 4% paraformaldehyde (PFA) in phosphate-buffered saline (4% PFA/PBS) for 10 minutes. Subsequently, the cells were incubated with the blocking buffer (PBS supplemented with 0.2 mM Ca<sup>2+</sup>, 2 mM Mg<sup>2+</sup>, and 0.5% bovine serum albumin) for 30 minutes, primary antibodies (10 µg/mL in the blocking buffer) for 1 hour, and Alexa Fluor 488-conjugated anti-mouse IgG (1:400 dilution in the blocking buffer) for 45 minutes.

For immunocytochemistry of NK cells, the suspension of NK cells was centrifuged at 270×g for 5 minutes at room temperature, and the cell pellet was suspended in and fixed

with 4% PFA/PBS for 10 minutes. The cells were further suspended in the blocking buffer for 30 minutes, primary antibodies (10 µg/mL in the blocking buffer) for 2 hours, and Alexa Fluor 488-conjugated anti-mouse IgG (1:400 dilution in the blocking buffer) for 45 minutes. 4',6-Diamidino-2-phenylindole (Thermo Fisher Scientific, Inc.) was used for nuclear staining of CHO-K1, CHO/hKLRG1, and NK cells. Fluorescence images were acquired using a 40× objective on a digital microscope (BZ-X800; Keyence, Osaka, Japan).

## Results

Our flow cytometric analysis revealed that CHO/hKLRG1 cells highly express hKLRG1 on the cell surface.<sup>57</sup> In this study, we applied KLMab-1 and recKLMab-1 in immunocytochemistry using CHO/hKLRG1 cells and found that KLMab-1 and recKLMab-1, but not buffer control, bound to CHO/hKLRG1 cells (Fig. 1A). In particular, hKLRG1 was strongly detected at the plasma membrane. Both mAbs did not react to CHO-K1 cells (Fig. 1B). A commercially available anti-hKLRG1 mAb (clone SA231A2) also bound to CHO/hKLRG1 cells, but not CHO-K1 cells (Fig. 1A, B). This result shows that KLMab-1 and recKLMab-1 recognize exogenous hKLRG1 in immunocytochemistry.

We previously showed that KLMab-1 detects endogenously expressing hKLRG1 in human NK cells in flow cytometry.<sup>57</sup> In this study, we incubated NK cells with KLMab-1 and recKLMab-1 and found that both mAbs, as well as SA231A2, bound to NK cells (Fig. 1C). This result demonstrates that KLMab-1 and recKLMab-1 recognize endogenous hKLRG1 in immunocytochemistry.

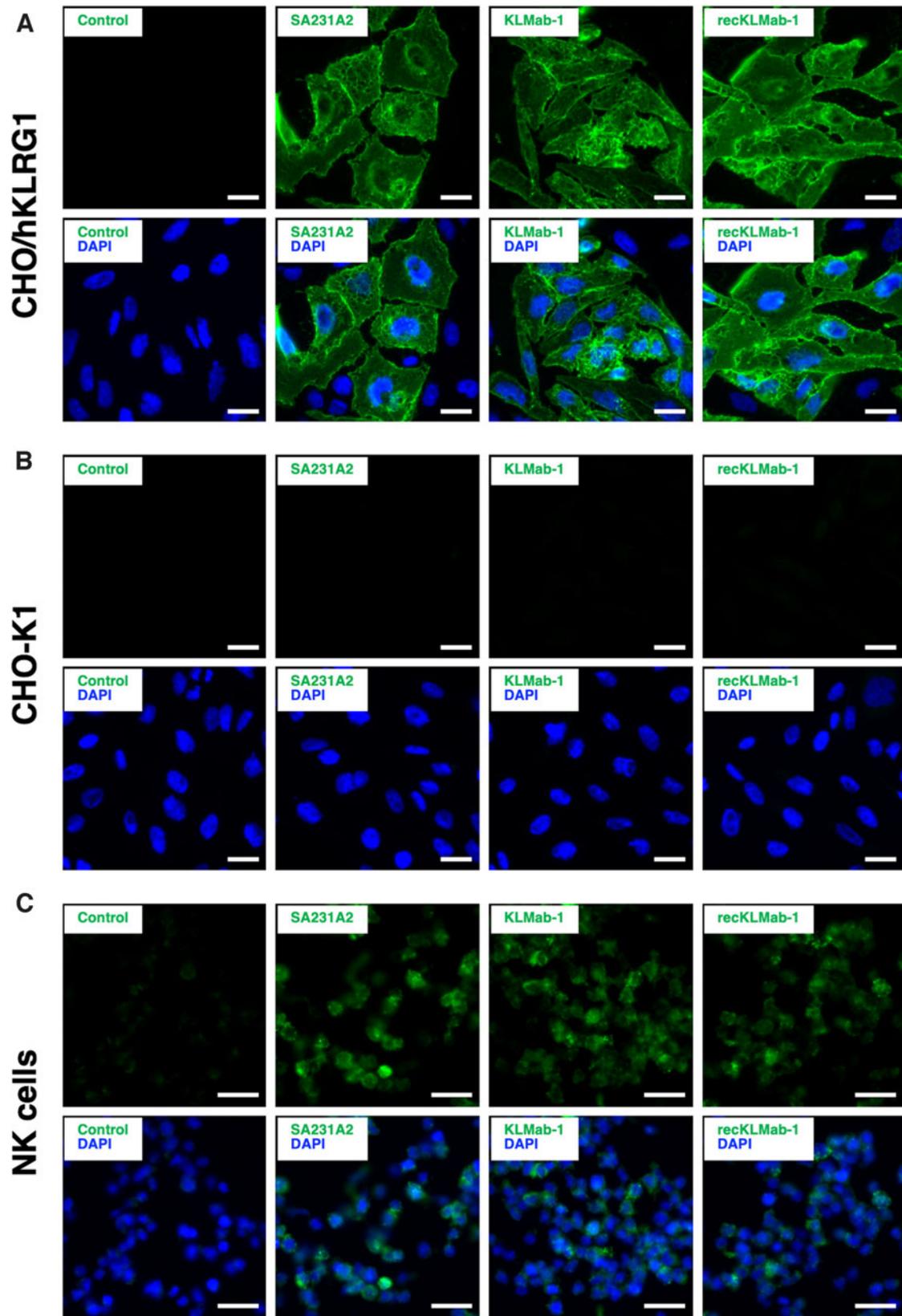
## Discussion

In this study, we demonstrated that KLMab-1 and recKLMab-1 were applicable for immunocytochemistry against endogenous and exogenous hKLRG1. The mAbs would become useful tools for the detection of hKLRG1-positive T and NK cells in viral infectious diseases and cancers.

KLMab-1 and recKLMab-1 provided images with high signal-to-noise ratios against not only exogenous hKLRG1 overexpressed in CHO-K1 cells but also endogenous hKLRG1 expressed in NK cells. Our previous study showed that KLMab-1 weakly detected endogenous hKLMab-1 in flow cytometry.<sup>57</sup> We suppose that KLMab-1 and recKLMab-1 are suitable for immunocytochemistry. Moreover, KLMab-1 and recKLMab-1 strongly detected hKLRG1 at the plasma membrane, which was clearly represented in CHO/hKLRG1 cells. The result indicates that the mAbs would be able to identify the intracellular distribution of hKLRG1, especially by colabeling the cells with any organelle markers. The information about the intracellular distribution of hKLRG1 would provide advantages in uncovering the unknown functions of KLRG1.

To uncover the KLRG1's roles and to detect KLRG1-positive cells, other applications of KLMab-1 and recKLMab-1 are also required. In the future, we would like to test other applications, including immunohistochemistry, immunoprecipitation, and Western blotting.

Furthermore, confirming the availability of KLMab-1 and recKLMab-1 in viral infectious diseases and cancers is one of our goals. Some studies have demonstrated that



**FIG. 1.** Immunocytochemistry of hKLRG1 using KLMab-1 and recKLMab-1. (A, B) CHO/hKLRG1 cells (A) or CHO-K1 (B) cells were incubated with buffer control, SA231A2 (10  $\mu\text{g}/\text{mL}$ ), KLMab-1 (10  $\mu\text{g}/\text{mL}$ ), or recKLMab-1 (10  $\mu\text{g}/\text{mL}$ ) for 1 hour. Subsequently, the cells were incubated with Alexa 488-conjugated anti-mouse IgG and DAPI for 45 minutes. (C) Immunocytochemistry of endogenously expressing hKLRG1. NK cells were incubated with SA231A2 (10  $\mu\text{g}/\text{mL}$ ), KLMab-1 (10  $\mu\text{g}/\text{mL}$ ), or recKLMab-1 (10  $\mu\text{g}/\text{mL}$ ) for 2 hours. Subsequently, NK cells were incubated with Alexa 488-conjugated anti-mouse IgG and DAPI for 45 minutes. Scale bars, 20  $\mu\text{m}$ . CHO, Chinese hamster ovary; DAPI, 4',6-diamidino-2-phenylindole; hKLRG1, human KLRG1; KLRG1, Killer cell lectin-like receptor subfamily G member 1; NK, natural killer.

anti-KLRG1 mAb reduces cytokine productions in KLRG1-overexpressed NK cells<sup>8</sup> and KLRG1<sup>+</sup> CD4<sup>+</sup> T cells.<sup>58</sup> An anti-KLRG1 mAb reduces the progression of breast cancer cells.<sup>59</sup> In addition, blocking of KLRG1 signaling in CD8<sup>+</sup> T cells by anti-E-cadherin antibody reduces the proliferation of the cells.<sup>60</sup> These studies would promote us to investigate the antitumor and/or antiviral effects of KLMab-1 and recKLMab-1.

#### Author Disclosure Statement

No competing financial interests exist.

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#### References

1. Kaech SM, Wherry EJ. Heterogeneity and cell-fate decisions in effector and memory CD8<sup>+</sup> T cell differentiation during viral infection. *Immunity* 2007;27:393–405; doi: 10.1016/j.immuni.2007.08.007
2. Huntington ND, Cursons J, Rautela J. The cancer-natural killer cell immunity cycle. *Nat Rev Cancer* 2020;20:437–454; doi: 10.1038/s41568-020-0272-z.
3. Bjorkstrom NK, Strunz B, Ljunggren HG. Natural killer cells in antiviral immunity. *Nat Rev Immunol* 2022;22:112–123; doi: 10.1038/s41577-021-00558-3
4. Thommen DS, Schumacher TN. T cell dysfunction in cancer. *Cancer Cell* 2018;33:547–562; doi: 10.1016/j.ccell.2018.03.012
5. Dyck L, Mills KHG. Immune checkpoints and their inhibition in cancer and infectious diseases. *Eur J Immunol* 2017;47:765–779; doi: 10.1002/eji.201646875
6. Ribas A, Wolchok JD. Cancer immunotherapy using checkpoint blockade. *Science* 2018;359:1350–1355; doi: 10.1126/science.aar4060
7. Kraehenbuehl L, Weng CH, Eghbali S, et al. Enhancing immunotherapy in cancer by targeting emerging immunomodulatory pathways. *Nat Rev Clin Oncol* 2022;19:37–50; doi: 10.1038/s41571-021-00552-7
8. Robbins SH, Nguyen KB, Takahashi N, et al. Cutting edge: Inhibitory functions of the killer cell lectin-like receptor G1 molecule during the activation of mouse NK cells. *J Immunol* 2002;168:2585–2589; doi: 10.4049/jimmunol.168.6.2585
9. Marcolino I, Przybylski GK, Koschella M, et al. Frequent expression of the natural killer cell receptor KLRG1 in human cord blood T cells: Correlation with replicative history. *Eur J Immunol* 2004;34:2672–2680; doi: 10.1002/eji.200425282
10. Guthmann MD, Tal M, Pecht I. A secretion inhibitory signal transduction molecule on mast cells is another C-type lectin. *Proc Natl Acad Sci U S A* 1995;92:9397–9401; doi: 10.1073/pnas.92.20.9397
11. Tourdot BE, Brenner MK, Keough KC, et al. Immunoreceptor tyrosine-based inhibitory motif (ITIM)-mediated inhibitory signaling is regulated by sequential phosphorylation mediated by distinct nonreceptor tyrosine kinases: A case study involving PECAM-1. *Biochemistry* 2013;52:2597–2608; doi: 10.1021/bi301461t
12. Tessmer MS, Fugere C, Stevenaert F, et al. KLRG1 binds cadherins and preferentially associates with SHIP-1. *Int Immunol* 2007;19:391–400; doi: 10.1093/intimm/dxm004
13. Herndler-Brandstetter D, Ishigame H, Shinnakasu R, et al. KLRG1<sup>+</sup> effector CD8<sup>+</sup> T cells lose KLRG1, differentiate into all memory T cell lineages, and convey enhanced protective immunity. *Immunity* 2018;48:716.e8–729.e8; doi: 10.1016/j.immuni.2018.03.015
14. Grundemann C, Bauer M, Schweier O, et al. Cutting edge: Identification of E-cadherin as a ligand for the murine killer cell lectin-like receptor G1. *J Immunol* 2006;176:1311–1315; doi: 10.4049/jimmunol.176.3.1311
15. Ito M, Maruyama T, Saito N, et al. Killer cell lectin-like receptor G1 binds three members of the classical cadherin family to inhibit NK cell cytotoxicity. *J Exp Med* 2006;203:289–295; doi: 10.1084/jem.20051986
16. Rosshart S, Hofmann M, Schweier O, et al. Interaction of KLRG1 with E-cadherin: New functional and structural insights. *Eur J Immunol* 2008;38:3354–3364; doi: 10.1002/eji.200838690
17. Saleh R, Taha RZ, Toor SM, et al. Expression of immune checkpoints and T cell exhaustion markers in early and advanced stages of colorectal cancer. *Cancer Immunol Immunother* 2020;69:1989–1999; doi: 10.1007/s00262-020-02593-w.
18. Guo Y, Feng Y, Fan P, et al. Expression and clinical significance of KLRG1 and 2B4 on T cells in the peripheral blood and tumour of patients with cervical cancer. *Immunol Invest* 2022;51(3):670–687; doi: 10.1080/08820139.2020.1867567
19. Asano T, Nanamiya R, Takei J, et al. Development of anti-mouse CC chemokine receptor 3 monoclonal antibodies for flow cytometry. *Monoclon Antib Immunodiagn Immunother* 2021;40:107–112; doi: 10.1089/mab.2021.0009
20. Asano T, Suzuki H, Tanaka T, et al. C<sub>3</sub>Mab-3: A monoclonal antibody for mouse CCR3 for flow cytometry monoclon. *Antib Immunodiagn Immunother* 2022;41:74–79; doi: 10.1089/mab.2021.0062
21. Saito M, Harigae Y, Li G, et al. C<sub>3</sub>Mab-2: An anti-mouse CCR3 monoclonal antibody for immunocytochemistry. *Monoclon Antib Immunodiagn Immunother* 2022;41:45–49; doi: 10.1089/mab.2021.0050
22. Tanaka T, Nanamiya R, Takei J, et al. Development of anti-mouse CC chemokine receptor 8 monoclonal antibodies for flow cytometry. *Monoclon Antib Immunodiagn Immunother* 2021;40:65–70; doi: 10.1089/mab.2021.0005
23. Saito M, Suzuki H, Tanaka T, et al. Development of an anti-mouse CCR8 monoclonal antibody (C<sub>8</sub>Mab-1) for flow cytometry and immunocytochemistry monoclon. *Antib Immunodiagn Immunother* 2022; [Online ahead of print]; doi: 10.1089/mab.2021.0069
24. Suzuki H, Saito M, Asano T, et al. C<sub>8</sub>Mab-3: An anti-mouse CCR8 monoclonal antibody for immunocytochemistry monoclon. *Antib Immunodiagn Immunother* 2022;41:110–114; doi: 10.1089/mab.2022.0002
25. Saito M, Tanaka T, Asano T, et al. C<sub>8</sub>Mab-2: An anti-mouse C–C motif chemokine receptor 8 monoclonal antibody for immunocytochemistry. *Monoclon Antib Immunodiagn Immunother* 2022;41:115–119; doi: 10.1089/mab.2021.0045

26. Nanamiya R, Takei J, Asano T, et al. Development of anti-human CC chemokine receptor 9 monoclonal antibodies for flow cytometry. *Monoclon Antib Immunodiagn Immunother* 2021;40:101–106; doi: 10.1089/mab.2021.0007
27. Saito M, Suzuki H, Harigae Y, et al. C<sub>9</sub>Mab-1: An anti-mouse CCR9 monoclonal antibody for immunocytochemistry. *Monoclon Antib Immunodiagn Immunother* 2022;41:120–124; doi: 10.1089/mab.2021.0052
28. Kawabata H, Ohishi T, Suzuki H, et al. A defucosylated mouse anti-CD10 monoclonal antibody (31-mG2a-f) exerts antitumor activity in a mouse xenograft model of renal cell cancers monoclon. *Antib Immunodiagn Immunother* 2022; [Online ahead of print], doi: 10.1089/mab.2021.0049
29. Kawabata H, Suzuki H, Ohishi T, et al. A defucosylated mouse anti-CD10 monoclonal antibody (31-mG2a-f) exerts antitumor activity in a mouse xenograft model of CD10-overexpressed tumors monoclon. *Antib Immunodiagn Immunother* 2022;41:59–66; doi: 10.1089/mab.2021.0048
30. Yamada S, Kaneko MK, Sayama Y, et al. Development of Novel Mouse Monoclonal Antibodies Against Human CD19. *Monoclon Antib Immunodiagn Immunother* 2020;39:45–50; doi: 10.1089/mab.2020.0003
31. Furusawa Y, Kaneko MK, Kato Y. Establishment of C<sub>20</sub>Mab-11, a novel anti-CD20 monoclonal antibody, for the detection of B cells. *Oncol Lett* 2020;20:1961–1967; doi: 10.3892/ol.2020.11753
32. Furusawa Y, Kaneko MK, Kato Y. Establishment of an anti-CD20 monoclonal antibody (C<sub>20</sub>Mab-60) for immunohistochemical analyses. *Monoclon Antib Immunodiagn Immunother* 2020;39:112–116; doi: 10.1089/mab.2020.0015
33. Yamada S, Itai S, Nakamura T, et al. Detection of high CD44 expression in oral cancers using the novel monoclonal antibody, C<sub>44</sub>Mab-5. *Biochem Biophys Rep* 2018;14:64–68; doi: 10.1016/j.bbrep.2018.03.007
34. Itai S, Fujii Y, Nakamura T, et al. Establishment of CMab-43, a sensitive and specific anti-CD133 monoclonal antibody, for immunohistochemistry. *Monoclon Antib Immunodiagn Immunother* 2017;36:231–235; doi: 10.1089/mab.2017.0031
35. Hosono H, Ohishi T, Takei J, et al. The anti-epithelial cell adhesion molecule (EpCAM) monoclonal antibody EpMab-16 exerts antitumor activity in a mouse model of colorectal adenocarcinoma. *Oncol Lett* 2020;20:383; doi: 10.3892/ol.2020.12246
36. Kaneko MK, Ohishi T, Takei J, et al. AntiEpCAM monoclonal antibody exerts antitumor activity against oral squamous cell carcinomas. *Oncol Rep* 2020;44:2517–2526; doi: 10.3892/or.2020.7808
37. Asano T, Ohishi T, Takei J, et al. Anti-HER3 monoclonal antibody exerts antitumor activity in a mouse model of colorectal adenocarcinoma. *Oncol Rep* 2021;46(2):173; doi: 10.3892/or.2021.8124
38. Yamada S, Itai S, Nakamura T, et al. Monoclonal antibody L<sub>1</sub>Mab-13 detected human PD-L1 in lung cancers. *Monoclon Antib Immunodiagn Immunother* 2018;37:110–115; doi: 10.1089/mab.2018.0004
39. Furusawa Y, Kaneko MK, Nakamura T, et al. Establishment of a monoclonal antibody PMab-231 for tiger podoplanin. *Monoclon Antib Immunodiagn Immunother* 2019;38:89–95; doi: 10.1089/mab.2019.0003
40. Furusawa Y, Takei J, Sayama Y, et al. Development of an anti-bear podoplanin monoclonal antibody PMab-247 for immunohistochemical analysis. *Biochem Biophys Rep* 2019;18:100644; doi: 10.1016/j.bbrep.2019.100644
41. Furusawa Y, Yamada S, Itai S, et al. Establishment of a monoclonal antibody PMab-233 for immunohistochemical analysis against Tasmanian devil podoplanin. *Biochem Biophys Rep* 2019;18:100631; doi: 10.1016/j.bbrep.2019.100631
42. Furusawa Y, Yamada S, Itai S, et al. PMab-219: A monoclonal antibody for the immunohistochemical analysis of horse podoplanin. *Biochem Biophys Rep* 2019;18:100616; doi: 10.1016/j.bbrep.2019.01.009
43. Furusawa Y, Yamada S, Itai S, et al. PMab-210: A monoclonal antibody against pig podoplanin. *Monoclon Antib Immunodiagn Immunother* 2019;38:30–36; doi: 10.1089/mab.2018.0038
44. Furusawa Y, Yamada S, Nakamura T, et al. PMab-235: A monoclonal antibody for immunohistochemical analysis against goat podoplanin. *Heliyon* 2019;5:e02063; doi: 10.1016/j.heliyon.2019.e02063
45. Kaneko MK, Sano M, Takei J, et al. Development and characterization of anti-sheep podoplanin monoclonal antibodies PMab-253 and PMab-260. *Monoclon Antib Immunodiagn Immunother* 2020;39:144–155; doi: 10.1089/mab.2020.0018
46. Kato Y, Furusawa Y, Itai S, et al. Establishment of an anticetacean podoplanin monoclonal antibody PMab-237 for immunohistochemical analysis. *Monoclon Antib Immunodiagn Immunother* 2019;38:108–113; doi: 10.1089/mab.2019.0013
47. Kato Y, Furusawa Y, Yamada S, et al. Establishment of a monoclonal antibody PMab-225 against alpaca podoplanin for immunohistochemical analyses. *Biochem Biophys Rep* 2019;18:100633; doi: 10.1016/j.bbrep.2019.100633
48. Kato Y, Yamada S, Furusawa Y, et al. PMab-213: A monoclonal antibody for immunohistochemical analysis against pig podoplanin. *Monoclon Antib Immunodiagn Immunother* 2019;38:18–24; doi: 10.1089/mab.2018.0048
49. Kato Y, Furusawa Y, Sano M, et al. Development of an anti-sheep podoplanin monoclonal antibody PMab-256 for immunohistochemical analysis of lymphatic endothelial cells. *Monoclon Antib Immunodiagn Immunother* 2020;39:82–90; doi: 10.1089/mab.2020.0005
50. Takei J, Yamada S, Konnai S, et al. PMab-241 specifically detects bear podoplanin of lymphatic endothelial cells in the lung of brown bear. *Monoclon Antib Immunodiagn Immunother* 2019;38:282–284; doi: 10.1089/mab.2019.0038
51. Tanaka T, Asano T, Sano M, et al. Development of monoclonal antibody PMab-269 against california sea lion podoplanin. *Monoclon Antib Immunodiagn Immunother* 2021;40:124–133; doi: 10.1089/mab.2021.0011
52. Goto N, Suzuki H, Tanaka T, et al. Development of a monoclonal antibody PMab-292 against ferret podoplanin. *Monoclon Antib Immunodiagn Immunother* 2022;41:101–109; doi: 10.1089/mab.2021.0067
53. Yamada S, Itai S, Nakamura T, et al. PMab-52: Specific and sensitive monoclonal antibody against cat podoplanin for immunohistochemistry. *Monoclon Antib Immunodiagn Immunother* 2017;36:224–230; doi: 10.1089/mab.2017.0027
54. Takei J, Asano T, Nanamiya R, et al. Development of anti-human T cell immunoreceptor with Ig and ITIM domains (TIGIT) monoclonal antibodies for flow cytometry. *Monoclon Antib Immunodiagn Immunother* 2021;40:71–75; doi: 10.1089/mab.2021.0006
55. Sayama Y, Kaneko MK, Kato Y. Development and characterization of TrMab6, a novel anti-TROP2 monoclonal

- antibody for antigen detection in breast cancer. *Mol Med Rep* 2021;23(2):92; doi: 10.3892/mmr.2020.11731
56. Sayama Y, Kaneko MK, Takei J, et al. Establishment of a novel anti-TROP2 monoclonal antibody TrMab-29 for immunohistochemical analysis. *Biochem Biophys Rep* 2021;25:100902; doi: 10.1016/j.bbrep.2020.100902
57. Asano T, Nanamiya R, Tanaka T, et al. Development of antihuman killer cell lectin-like receptor subfamily G member 1 monoclonal antibodies for flow cytometry. *Monoclon Antib Immunodiagn Immunother* 2021;40:76–80; doi: 10.1089/mab.2021.0008
58. Hu Z, Zhao HM, Li CL, et al. The role of KLRG1 in human CD4<sup>+</sup> T-cell immunity against tuberculosis. *J Infect Dis* 2018;217:1491–1503; doi: 10.1093/infdis/jiy046
59. Chan IS, Knutsdottir H, Ramakrishnan G, et al. Cancer cells educate natural killer cells to a metastasis-promoting cell state. *J Cell Biol* 2020;219(9):e202001134; doi: 10.1083/jcb.202001134
60. Henson SM, Franzese O, Macaulay R, et al. KLRG1 signaling induces defective Akt (ser473) phosphorylation and proliferative dysfunction of highly differentiated CD8<sup>+</sup> T cells. *Blood* 2009;113:6619–6628; doi: 10.1182/blood-2009-01-199588

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